

# Supplements

## **Chiral analysis of glycerol phosphates - Can bacteria biosynthesize heterochiral phospholipid membranes?**

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### Detection of heterochirality in lipid membranes

Two commercially available standards (1,2-dimyristoyl-sn-glycero-3-phosphocholine and 2,3-dimyristoyl-sn-glycero-1-phosphocholine) were added in HC2Cl2-MeOH solution to the lyophilized cells; lipids were extracted and the enantiomers was found. It was shown that the second enantiomer was not present after extraction and chiral HPLC, the respective proportion being below 1% of total enantiomers. Chiral HPLC of both standards was also performed, and it was found that the second enantiomer was not present in the commercial preparation, or its concentration was below 1%. This means that no phosphoryl group migration occurred. The areas of the ion peaks at  $m/z$  734.5136 were compared. The same procedure was carried out with 1-oleoyl-2-palmitoyl-sn-glycero-3-phosphocholine and, in another experiment, with 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine; the detection was performed at  $m/z$  818.5919. This experiment proved that there was no acyl(s) migration. Finally, we may conclude that this approach proved that migration of neither the acyl(s) nor phosphoryl group took place in both, the mobile phase and the cells.

Table S1. Contents of G3P and G1P

| microorganism                         | G3P                | G1P            | reference  | ratio G3P/G1P |
|---------------------------------------|--------------------|----------------|------------|---------------|
| <i>Bacillus amyloliquefacience</i>    | 34                 | 11.5           | this paper | 3.0           |
| <i>Bacillus subtilis</i>              | 33                 | 10.9           | this paper | 3.0           |
| <i>Clavibacter michiganensis</i>      | 23                 | 5.3            | this paper | 4.3           |
| <i>Geobacillus stearothermophilus</i> | 25                 | 6.2            | this paper | 4.0           |
| <i>Saccharomyces pastorianus</i>      | 30                 | 0.0            | this paper | -             |
| <i>Escherichia coli</i>               | 49                 | ? <sup>b</sup> | A35a       | -             |
| <i>Spathaspora arborariae</i>         | 44/15 <sup>c</sup> | ?              | A35b       | -             |

<sup>a</sup> The values (units) are in  $\mu\text{M/L}$ .

<sup>b</sup> enantiomer analysis was not performed

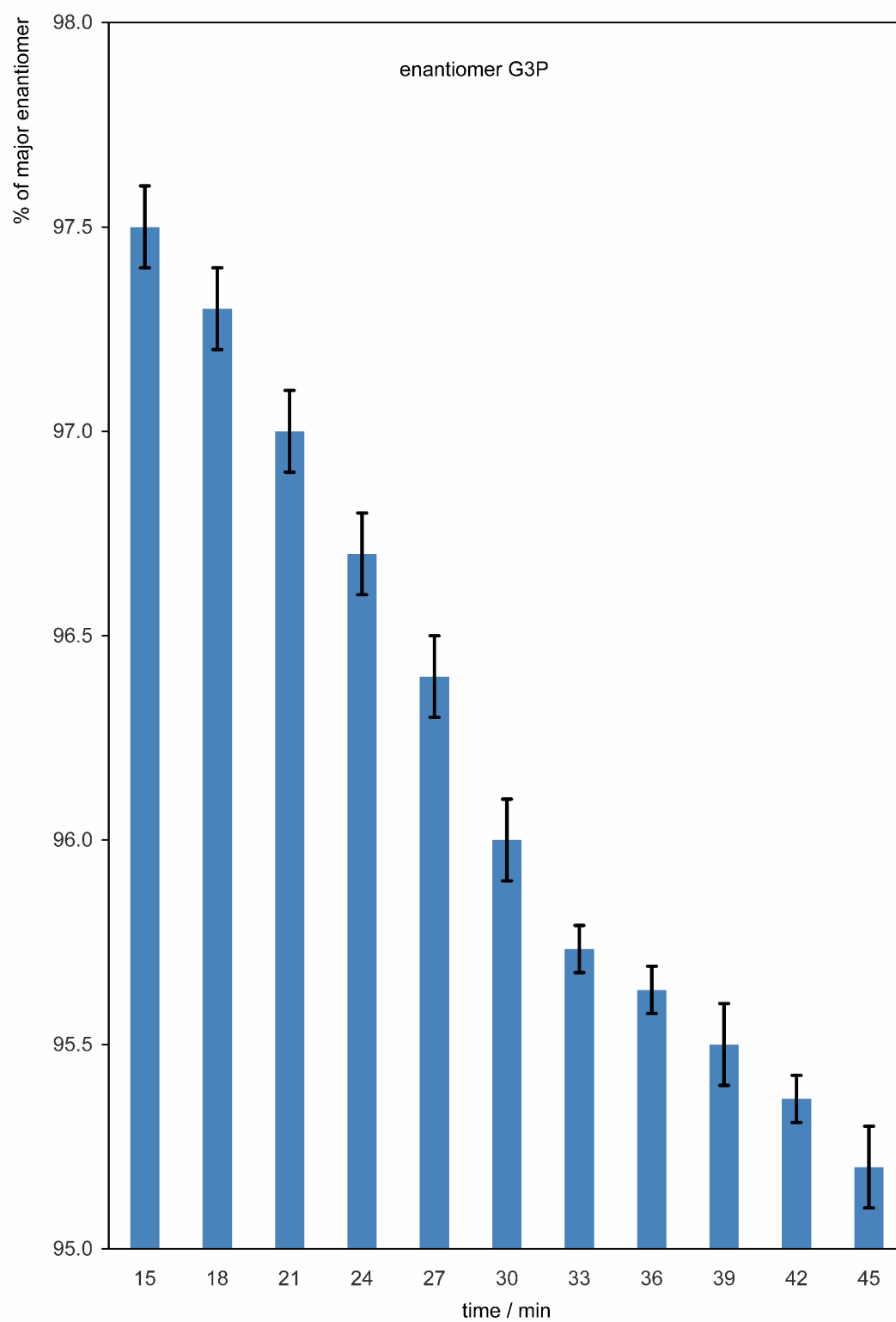
<sup>c</sup> aerobic cultivation/oxygen-limited cultivation

Table S2. Occurrence of enzymes in bacteria - glycerol-1-phosphate dehydrogenase.

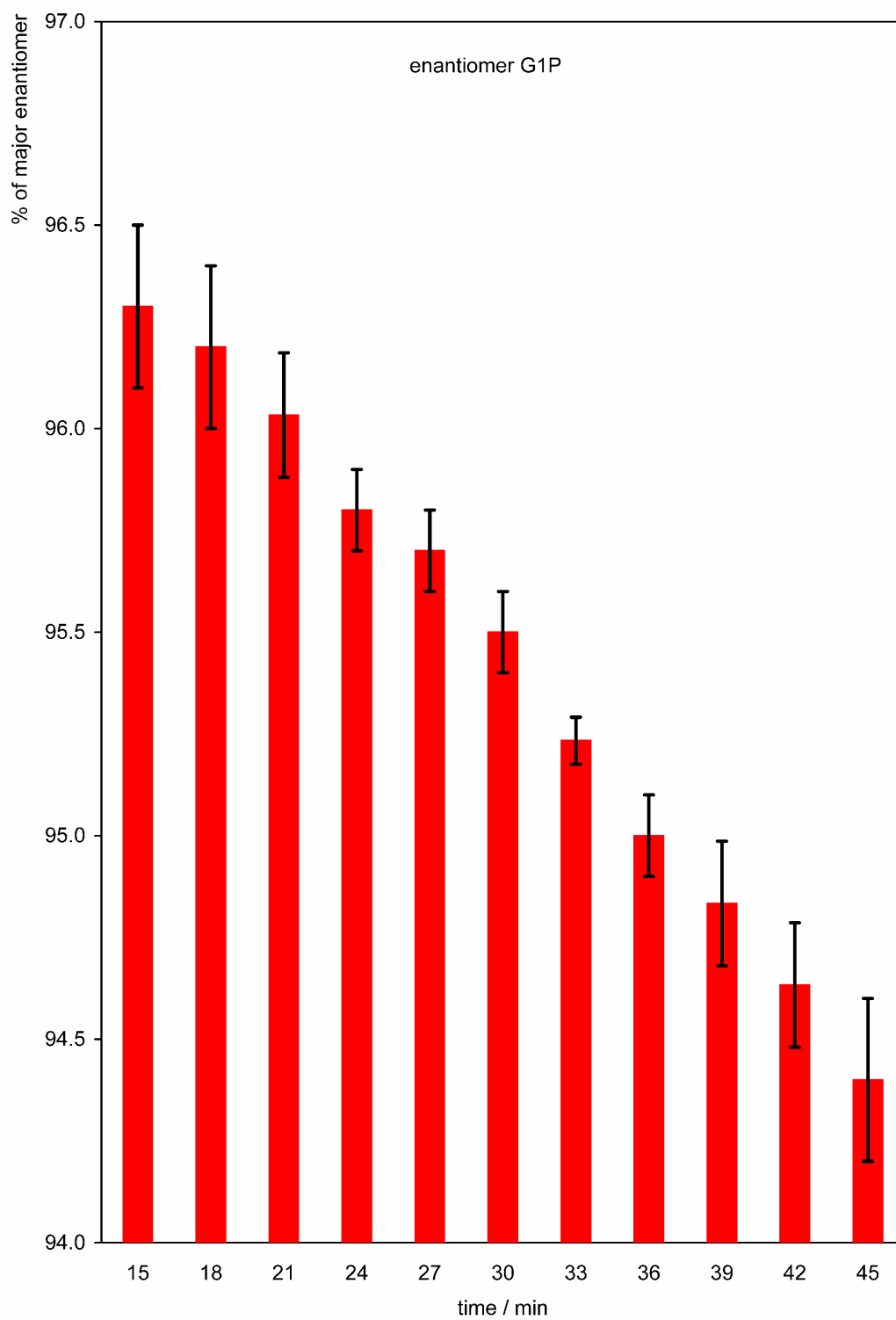
| bacterium                             | GenBank    |
|---------------------------------------|------------|
| <i>Bacillus amyloliquefaciens</i>     | KYC93433.1 |
| <i>Bacillus subtilis</i>              | AIW38374.1 |
| <i>Clavibacter</i> sp.                | GBL23066.1 |
| <i>Geobacillus stearothermophilus</i> | KFX35283.1 |

Table S3. Occurrence of enzymes in bacteria - glycerol-3-phosphate dehydrogenase.

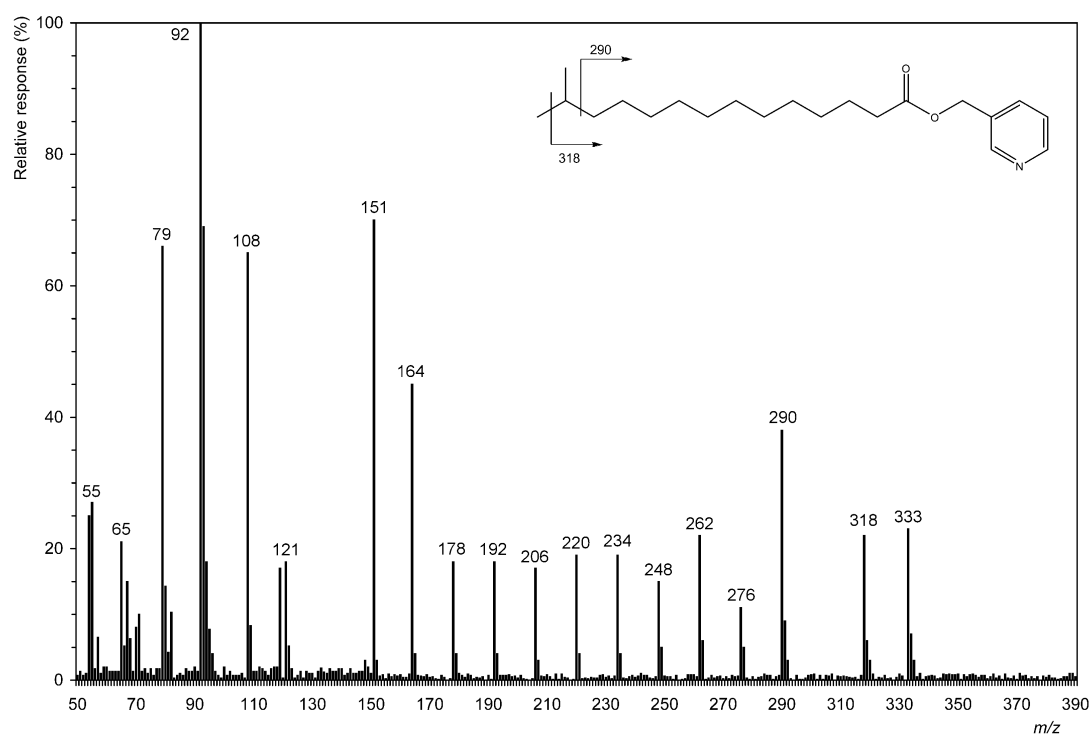
| bacterium                                                   | GenBank    |
|-------------------------------------------------------------|------------|
| <i>Bacillus amyloliquefaciens</i>                           | KYC86943.1 |
| <i>Bacillus subtilis</i>                                    | KOS72728.1 |
| <i>Clavibacter michiganensis</i> subsp. <i>nebraskensis</i> | OAH22261.1 |
| <i>Geobacillus stearothermophilus</i>                       | KYD34837.1 |



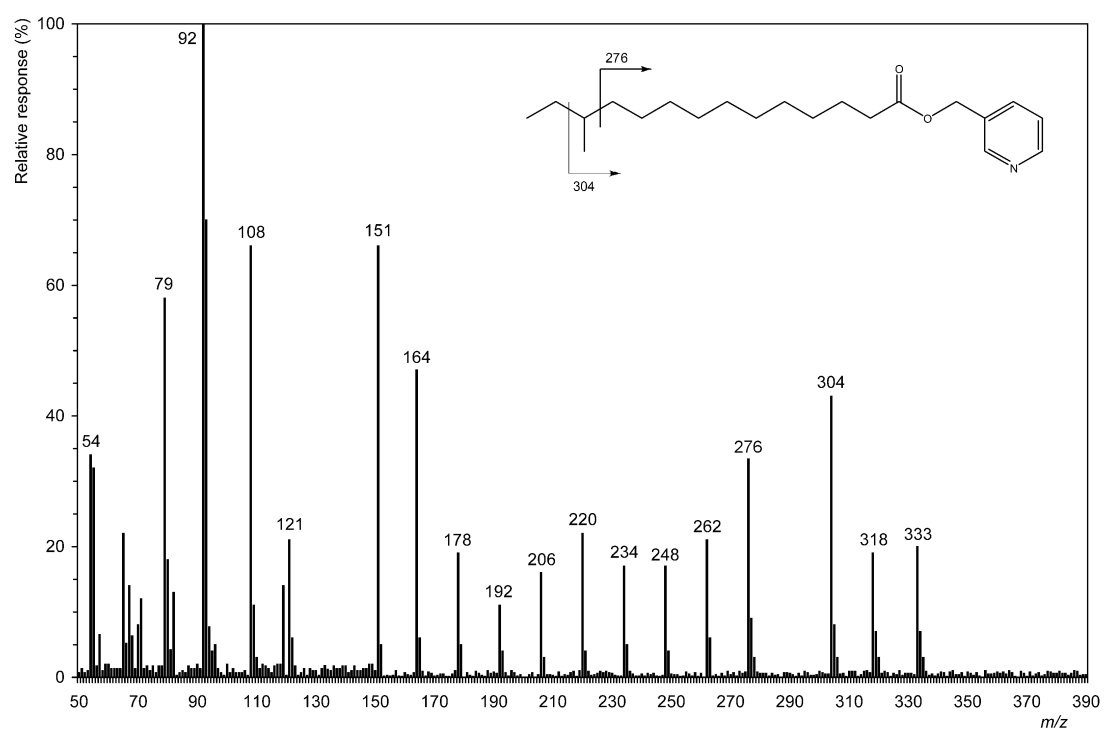
**Fig. 1S.** Time course of conversion of G3P into a racemic mixture.



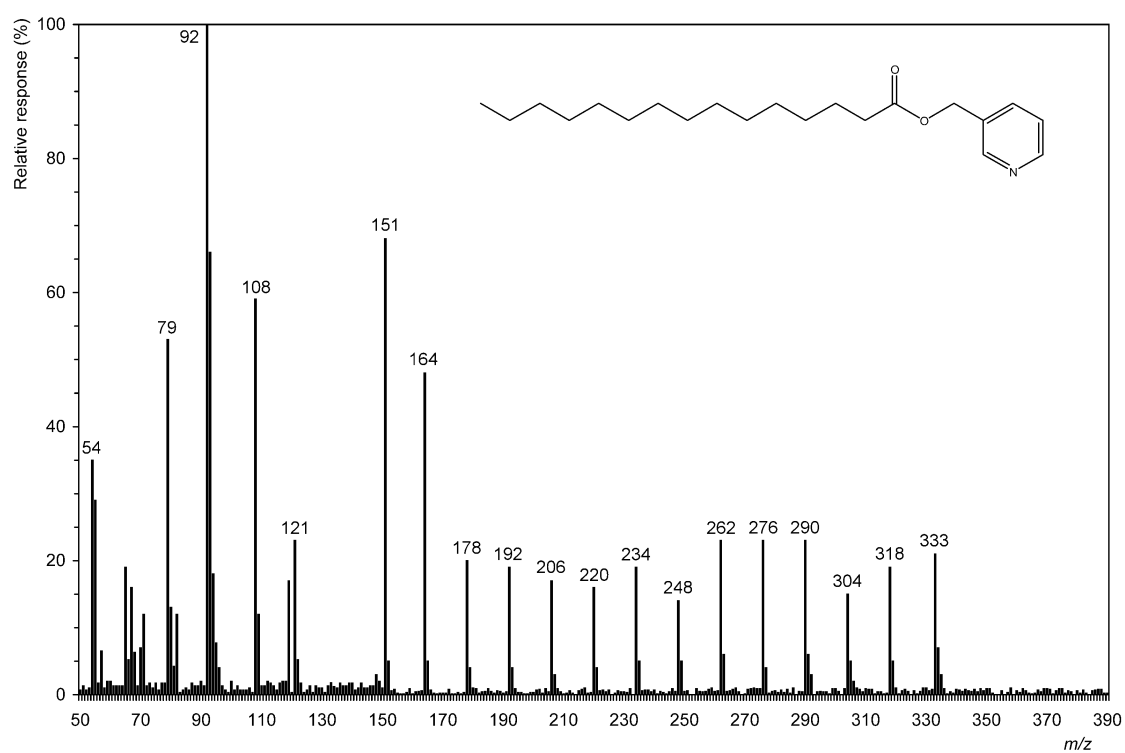
**Fig. 2S.** Time course of conversion of G1P into a racemic mixture.



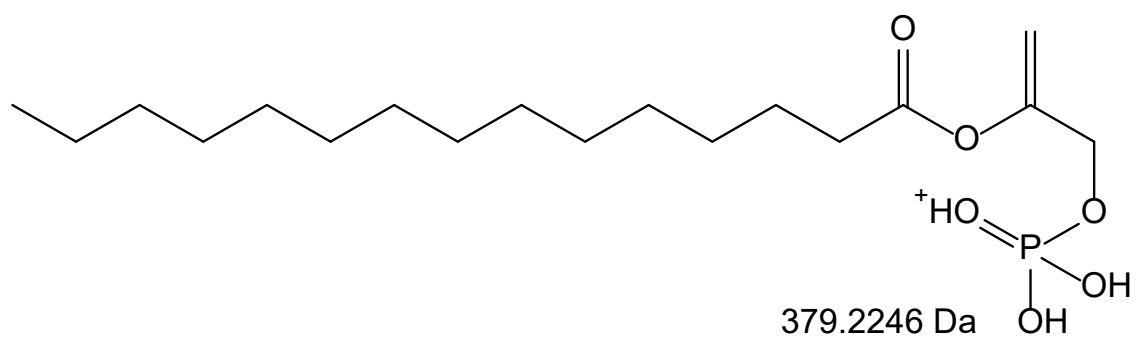
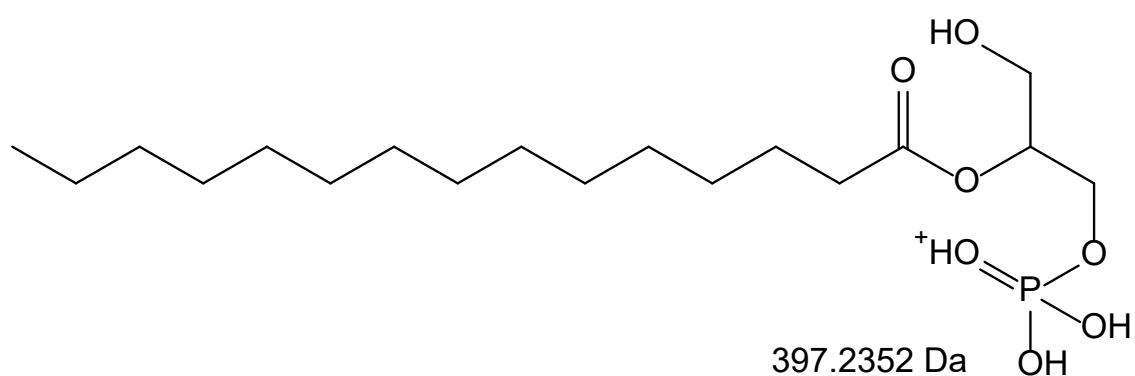
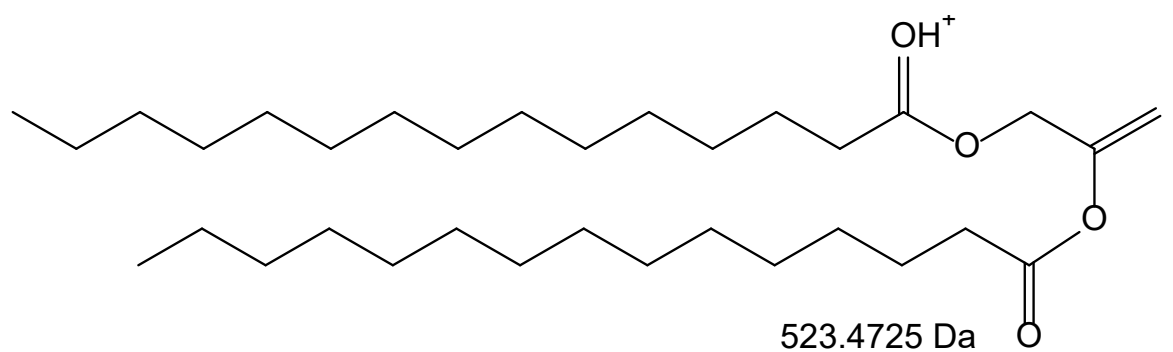
**Fig. 3S.** Mass spectrum of 3-pyridylcarbinol esters of 13-methyltetradecanoate.



**Fig. 4S.** Mass spectrum of 3-pyridylcarbinol esters of 12-methyltetradecanoate.



**Fig. 5S.** Mass spectrum of 3-pyridylcarbinol esters of pentadecanoate.



**Fig. 6S.** Molecular species 15:0/15:0-BMP provides fragment ions at  $m/z$  397.2352 and  $m/z$  379.2246; corresponding to FA chain loss and isomeric molecular species 15:0/15:0-PG produced a protonated fragment ion at  $m/z$  523.4725.